

A standardized nomenclature for the rods and cones of the vertebrate retina

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SUMMARY. We propose a standardized naming system for vertebrate visual photoreceptors (i.e., rods and cones) that reflects our current understanding of their evolutionary history. Vertebrate photoreceptors have been studied for well over a century, but a fixed nomenclature for referring to orthologous cell types across diverse species has been lacking. Instead, photoreceptors have been variably - and often confusingly - named according to morphology, presence/absence of 'rhodopsin,' spectral sensitivity, chromophore usage, and/or the gene family of the opsin(s) they express. Here, we propose a unified nomenclature for vertebrate rods and cones that aligns with the naming systems of other retinal cell classes and that is based on the photoreceptor's putative ancestral derivation. This classification is informed by the functional, anatomical, developmental and molecular identities of the neuron as a whole, including the expression of deeply conserved transcription factors required for development. The proposed names will be applicable across all vertebrates and indicative of the widest-possible range of properties, including their postsynaptic wiring, and hence will allude to their common and species-specific roles in vision. Furthermore, the naming system is open-ended to accommodate the future discovery of as-yet unknown photoreceptor types.

Figure 1. Summary of proposed naming system for vertebrate photoreceptors and its relationship to prior systems¹⁻⁴.

New name	Common names	Morphological names	Typical opsin
Rod/ P0	Rod	Rod	RH1
P1	Red/L	Ancestral / single cones Fish twin/ double/triple	LWS
P2	Green/M		RH2
P3	Blue/S		SWS2
P4	Violet/UV/S		SWS1
P5	Principal	Tetrapod double cones	LWS
P6	Accessory		LWS
P7	'Green-rod'	'Second rod' (amphibians)	SWS2

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INTRODUCTION. The common ancestor of extant vertebrates likely possessed a photoreceptor system consisting of one rod and four cone types, based on the shared presence of these five cell types in present-day jawed and jawless vertebrates^{5,6}. Accordingly, we begin our classification with these five types (Fig. 1). Following prior studies of other retinal cell classes, we propose a simple naming system in which the photoreceptor types are designated with a single letter ('P' for 'photoreceptor') followed by a number (Fig. 1). Thus, cones derived from the four ancestral single cone types will be designated P1-P4 (alternative: 'Type 1-4 photoreceptors'), and rods will be P0. Prior studies have shown that retinal cell types typically occur in regular spatial mosaics, in which cells are tiled in a non-random distribution with minimal spacing between neurons of the same type. A prior study of cone photoreceptor distributions in chicken suggested that mosaic spacing is also a feature of photoreceptor types⁷. Accordingly, we propose mosaic spacing as a working definition of photoreceptor type. Under this definition, regional variation in photoreceptor morphology or gene expression can occur in a single photoreceptor type, without needing to designate regional variants (e.g., foveal vs. peripheral cones in human) as distinct cell types. Our classification also includes the tetrapod double cone, which consists of principal (P5) and accessory (P6) members, and the so-called 'green rod' of amphibians (P7). The two members of the double cone likely arose in the common ancestor of tetrapods, whereas the 'green rod' appears to have arisen early in the amphibian lineage⁸. According to this system, zebrafish retinas possess P0-P4⁹, while chicken retinas have P0-P6¹⁰. Eutherian mammals, which include mice and humans, have P0, P1, and P4 types¹¹. Depending on context, special notation may be required to distinguish unique subtypes of photoreceptors, such as human 'green/M' and 'red/L' cones, which differ only in the expression of paralogous LWS opsin genes. In this case, we propose the designations P1-M and P1-L, for 'M' and 'L' cones, respectively. Additional photoreceptor types can readily be added to this system when they are discovered.

91

THE PROBLEM

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Current naming systems for vertebrate ciliary photoreceptors represent a grab bag of species-specific schemes, typically based on a single functional, morphological, or molecular feature. For example, the cone types of the human eye are often referred to as red/L, green/M, and blue/S. This naming system references the wavelength of cones' maximal spectral sensitivity as imparted by the visual pigment^{1,12} (opsin + 11-cis retinal chromophore), and oil droplet filtering¹³, where present. Since the 1990's¹⁴⁻¹⁶ vertebrate visual photoreceptors have commonly been classified by the main opsin photopigment that they express: RH1 (rods) and LWS, RH2, SWS2, SWS1 (cones). These ancient gene sub-families were already present in the last common vertebrate ancestor, and exhibit remarkable evolutionary conservation, for example in the spectral ranges of the visual pigments, and, as we explain here, in the cell types in which they are expressed. Even so,

105 naming cone types by their photopigment can be problematic. For example,
106 human red and green cones differ by a single feature, namely which of two
107 paralogues of the LWS opsin gene is expressed¹¹. Human L and M cones
108 derive from a single ancestral photoreceptor type, and homologs of this cell
109 type are present in the eyes of most extant vertebrates⁴. Why, then, should
110 this photoreceptor type not share a common designation across species?

111 In general, the names of photoreceptor types fall into two major groups:
112 those that refer to spectral properties and/or the expressed type(s) of opsin
113 (Box 1), and those that describe morphological properties, such as their size
114 or linkage association with other photoreceptor types (Box 2). However,
115 'spectral' ('red/L') and opsin ('LWS') terms are problematic because the
116 properties that they reference are subject to routine variation within and
117 between species⁴. Likewise, morphological terms such as 'single', 'double',
118 'twin', and 'paired' mean different things in different animals³: In fishes, the
119 latter three refer to coupled combinations of ancestral 'red' and 'green'
120 cones¹⁷, commonly found as part of mosaics of variable regularity^{18,19}. In
121 contrast, 'tetrapod double cones'²⁰ are molecularly^{17,21}, morphologically^{7,22,23}
122 and functionally¹³ distinct from the four ancestral single cones²⁴ and likely
123 have a distinct evolutionary origin.

124 To overcome such naming confusions, current progress in life sciences
125 requires cell-type definitions that encompass a wide spectrum of features^{25–}
126 ²⁷, ideally combining morphological²⁸, functional²⁹, developmental^{17,30} and
127 molecular traits³¹. We also emphasize the importance of deeply conserved
128 transcription factors as type-defining features in our classification.

129 The vertebrate retina, with its planar structure and regularly tiled architecture
130 has long served as a central model system for cell type taxonomy. Retinal
131 neurons form spatially repeating units³² that anchor efforts to define types
132 and their plausible subdivisions across animal brains. In some species such
133 as the mouse, the catalogue of retinal neuron types is probably close to
134 complete. This momentous achievement has been gradually unlocked by
135 the use of transgenic tools³³, large-scale physiological recording
136 techniques^{29,34}, EM-connectomics^{28,35,36}, and, perhaps most importantly
137 single cell transcriptomics^{26,27,31,37–39}, which can assign a genome-wide
138 molecular signature to anatomical and functional catalogues.

139 The bipolar cells of the mouse retina were amongst the first to be 'solved' in
140 this way^{36,37,40}, and it has taken only a few more years for the tabulation of
141 other retinal neuron types in a number of species to become increasingly
142 complete. The mouse retina comprises ~130 neuron types: the rod, 2 cones,
143 1 horizontal cell, 15 bipolar cells^{35–37,41}, ~63 amacrine cells³⁸ and ~45
144 ganglion cells^{26,27,29}. Of these, all except the rods and cones are primarily
145 known by a numbered identity (e.g. H1 horizontal cell, Type 1 bipolar cell,
146 etc.). While aspects of these catalogues still require consensus and
147 alignment across species, in principle a numerical system gives each type
148 of neuron a unique and unmistakable identity. For example, a horizontal cell
149 that is orthologous to the H1 of mice⁴² also exists in lampreys⁴³,

150 zebrafish^{39,44}, chicken^{10,21,45} and humans^{39,46}. This understanding allows us
151 to make powerful inferences across the vertebrate tree of life.

152 Recent transcriptomic studies (e.g. Refs^{17,30,47,48}) investigated molecular
153 relationships of vertebrate photoreceptors across a broad range of species.
154 Among other findings, this work provided a molecular confirmation of what
155 had long been suspected⁴: Many vertebrate photoreceptor types appear to
156 be orthologous across multiple evolutionarily distant species. We believe
157 that the time has come to inscribe this knowledge into the names of
158 photoreceptor types.

159 **OUR PROPOSAL**

160 As a clear and systematic nomenclature for vertebrate photoreceptors, we
161 suggest a numbering scheme that simultaneously mirrors the cell's ancestry,
162 their typical relative abundance in the eye, their development, and their
163 systematically distinct postsynaptic wiring patterns (Box 3).

164 Ancestral red cones⁴, renamed P1, are probably the least derived type of
165 cone. In most non-avian vertebrate eyes, barring rods, P1 are the most
166 abundant, and probably also most important type of photoreceptor^{4,20,49}. P1
167 typically, but not always, expresses LWS-opsin, and the only sighted
168 vertebrates thought to lack P1 are rod-only species that typically live in
169 extreme low-light conditions. Human daylight vision is overwhelmingly driven
170 by P1 cones⁵⁰, which include both their 'L' and 'M' variants¹¹. In mouse, P1
171 includes both the 'green' sensitive dorsal and 'UV-sensitive' ventral LWS
172 cones⁵¹⁻⁵³.

173 Beyond P1, the other three 'original' single cones^{4,6} (ancestral green/RH2,
174 blue/SWS2, UV/SWS1) become P2-4, respectively. This order mirrors their
175 typical spectral order² (cf. Box 1), and their transcriptomic relatedness to
176 P1¹⁷, their anatomical wiring order in the retina ('spectral block wiring'^{11,9}), as
177 well as the order of their typical numerical abundance and relative sizes
178 ($P1 \geq P2 \geq P3 \geq P4$, e.g. chicken⁷, adult zebrafish⁵⁴, see also Box 2). Moreover,
179 this sequence mirrors the numerical order of cones' postsynaptic targets,
180 namely horizontal cells and bipolar cells. For example, the zebrafish H1
181 horizontal cell is the only one that connects with P1 cones, alongside P2,3⁴⁴.
182 H2 then links with P2-4 cones, while H3 links to P3,4. Likewise, the Type 1
183 bipolar cell of mice is the only cone-bipolar cell that preferentially targets P1
184 cones³⁶. The numerical system also elucidates evolutionary loss of cone
185 types, e.g. for eutherian mammals who retain only P1 and P4⁴.

186 After these original five photoreceptor types, the next most widespread
187 photoreceptor type in vertebrates is the tetrapod double cone^{20,23,24}, and we
188 suggest that this becomes P5,6 for the principal and accessory members,
189 respectively. This system also mirrors their likely evolutionary order of
190 appearance²⁰. We posit that the double cone should receive two numbers (5
191 and 6) rather than one because (i) it consists of two neurons rather than
192 one^{17,21,23}, (ii) each member makes independent connections to postsynaptic
193 targets²², and (iii), the two members almost certainly differ in their

194 physiological properties and roles in vision^{13,24,55}; and (iv) the two cell types
195 are morphologically²² and molecularly^{17,30} distinct.

196 Beyond these established photoreceptor types (see below), many
197 amphibians have a 'second-rod' type known traditionally as the 'second' or
198 'green rod' based on its microscopic appearance (and in contrast with the
199 canonical or 'red rod')⁵⁶⁻⁵⁸. Tentatively designated P7, the ancestry of this
200 eighth type of photoreceptor remains unresolved. However, its exclusive
201 presence in amphibians strongly suggests that unlike P5/6, P7 may have
202 emerged after present-day amphibians diverged from all other tetrapods.
203 Beyond P7, further hints of yet to be defined photoreceptor types include a
204 possible 'extra' type in some marsupials such as dunnarts⁵⁹ and subsets of
205 photoreceptors found in some snakes⁶⁰ and geckos⁶¹⁻⁶³. We posit that these,
206 or others, should be added onto the end of the proposed scheme if and when
207 appropriate.

208 Finally, we propose that rods be designated P0. Despite rods' substantial
209 molecular differentiation from cones¹⁷, both rods and cones share a common
210 ancestry, and the presence of rods (P0) and ancestral single cones (P1-4)
211 in cyclostomes^{6,43,64} strongly suggests that both were present in stem
212 vertebrates. While some species feature additional rod-like photoreceptors,
213 it remains unclear which, if any, represent genuine new types (see Box 3,
214 Definitions).

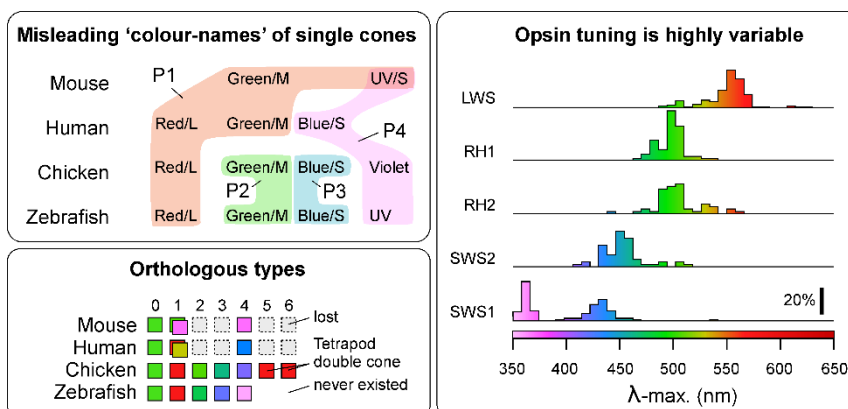
215 **A FINAL WORD**

216 We acknowledge that some common names for cones are deeply
217 engrained, both in the scientific literature but also in popular culture. For
218 example, the terms of 'L', 'M' and 'S' cones of the human eye, although
219 potentially misleading (discussed in Ref⁴), are unlikely to go away anytime
220 soon, or perhaps ever. However, as a community we can endeavour to
221 define the terms that we use in our communications by the simple and
222 hopefully uncontroversial numbering scheme suggested here. Perhaps, in
223 this way, it will eventually become more widely adopted.

224 **Box 1. Opsins and their spectral properties are poor indicators of cone identity**

225 Comparison of humans, zebrafish, and mice illustrates the central problem. Humans and
 226 zebrafish have 'red/L', 'green/M' and 'blue/S' cones, while zebrafish additionally have
 227 ultraviolet (UV) cones⁴, but human 'green/M' and 'blue/S' cones are evolutionarily unrelated
 228 to zebrafish 'green/M' and 'blue/S' cones¹⁷. The opsins of human 'green/M' and 'red/L' cones
 229 are orthologous to zebrafish 'red/L' (all express LWS), and human 'blue/S' to zebrafish UV
 230 (both express SWS1). However, this match by opsins is fortuitous in the sense that both
 231 human and zebrafish cones, where present, consistently express variants of their ancestrally
 232 linked opsins: LWS, RH2, SWS2 and SWS1 for P1-4, respectively². By contrast, the opsin
 233 scheme falls apart in mice because mouse P1 cones (which are often referred to as 'green/M'
 234 in reference to their 'green-shifted' LWS opsin) co-express the ancestral UV-opsin SWS1 in
 235 the ventral retina⁵¹⁻⁵³. The same ancestral neuron type P1 therefore transitions from
 236 'functionally green' to 'functionally UV' along the dorsal-ventral axis of the retina. Moreover
 237 mice retain the ancestral UV-cone P4, which like the SWS1-coexpressing P1 cones are more
 238 concentrated in the ventral retina⁶⁵. Mice therefore have two types of UV-sensitive cones in
 239 direct proximity. Similarly, some fish species including cichlids and salmonids are known to
 240 switch opsin expression in individual cones, such that P1 cones may express LWS or RH2
 241 opsins, and P4 cones may express SWS1 or SWS2 opsins depending on developmental stage
 242 or environmental cues⁶⁶⁻⁶⁸. While these examples illustrate the problem, they are not outliers
 243 in the vertebrate tree of life. The identity and wavelength specificity of expressed cone opsins
 244 is subject to routine variation^{1,2}, both across species (e.g. Ref⁶⁹) as well as within species
 245 (including by retinal region^{51,52}, life stage⁷⁰, season⁷¹⁻⁷³, and environment⁷⁴). It further depends
 246 on an opsin's associated chromophore (A₁ or A₂)^{75,76}, which also varies seasonably and
 247 according to life stage. In fact, opsins and their properties are an evolutionary hotspot, varying
 248 as species enter new visual niches^{2,77}. The identity or functional properties of opsins therefore
 249 do not reliably specify the identity of the neuron that expresses them.

250 A second issue is that a definition by 'colour' implies that wavelength selectivity is the only
 251 important characteristic of a photoreceptor. This is misleading⁴, because beyond wavelength
 252 selectivity, cone types systematically differ in their basic cellular physiology including their
 253 spatio-temporal properties⁷⁸⁻⁸⁰, as well as in their developmental postsynaptic wiring^{22,36,81} –
 254 all of which directly feed into their distinct roles in vision⁴⁹.



263 **Figure:** Left, Cone 'colour names' (top) do not necessarily align with orthologous neuron types across vertebrates
 264 (bottom, based on Ref⁴). Right, Sequence changes within vertebrate opsins can give rise to diverse spectral
 265 properties (based on Ref²). Note e.g. that a λ-max of ~500 nm is readily achieved by four out of five opsin families.
 266 Colour shadings as defined on the bottom right, by most common expressed opsin variant, independent of oil
 267 droplets, where present.

268 **Box 2. Cellular morphology is an imperfect indicator of photoreceptor identity**

269 Morphological definitions of photoreceptor types based on the shape of the outer segment
 270 (i.e., ‘rod’ vs. ‘cone’), association with other photoreceptors (e.g., ‘single’ vs. ‘double’ cones)
 271 or other cellular features are as problematic as opsin-based definitions. Photoreceptors can
 272 be grouped into ‘morphological types’, namely ‘single cones’ which tend to occur in isolation,
 273 ‘twin’ cones³ which comprise pairs made up of morphologically identical partners, and
 274 ‘double/triple cones’ consisting of asymmetric groups, often with ‘principal’ and ‘accessory’
 275 members^{3,7,24,82}. Single cones are occasionally further identified by other descriptors such as
 276 ‘long’, ‘short’⁸³, and miniature⁸⁴ in reference to their size and/or vertical location in the outer
 277 retina. However, there are many factors that influence the anatomical arrangement of
 278 photoreceptors, and like opsin or spectral identity, none are reliably stable across species, or
 279 within.

280 Zebrafish and chicken serve as two well studied examples. Adult zebrafish have a crystalline
 281 photoreceptor mosaic with P1-4 cones arranged at a fixed 2:2:1:1 stoichiometry in the adult^{9,85}.
 282 These six neurons are arranged in a tight lattice whereby P1,2 pairs alternate rows with C3
 283 and C4. Fishes have different patterns of C1-4 cones that vary in spectral content and
 284 regularity depending on developmental stage and retinal location. C1,2 pairs have been
 285 generally referred to as ‘paired cones’. If the two members have different morphology (usually
 286 C1>C2) or visual pigment content, they have been called ‘double cones’. If they appear
 287 morphologically equal and contain the same visual pigment, they have been termed ‘twin
 288 cones’⁸⁶.

289 In non-tetrapod vertebrates, all the above terms describe subsets of the same four ancestral
 290 cone types¹⁷, P1-4. One way to ascertain this is during development: In zebrafish, the retina
 291 does not start out in a crystalline arrangement that forces some cone types into pairs. Instead,
 292 larval cones are arranged independently^{44,85,87}. This developmental history is inscribed into the
 293 adult retina as the ‘larval patch’ near the optic disc, which never rearranges^{85,88}. Beyond this
 294 developmental evidence, the conclusion that all (zebra)fish cones are subsets of the same
 295 four ancestral ‘single’ cones is overwhelmingly supported by single cell transcriptomics^{17,89}. By
 296 contrast, many tetrapods, with the notable exception of eutherian mammals, do have an ‘extra’
 297 pair of cone-photoreceptors, called ‘the tetrapod double cone’²⁰ (P5,6). Unlike fish
 298 ‘double/twin/paired’ cones, this cone pair exists in parallel to P1,2. Birds and most reptiles

therefore have six ancestrally distinct types of cones. The four ‘original’ cones P1-4 plus the two members of the tetrapod double cone P5 and P6. This view is supported by extensive morphological^{7,13,22,23}, behavioural^{24,90} and molecular evidence^{17,21,91}.

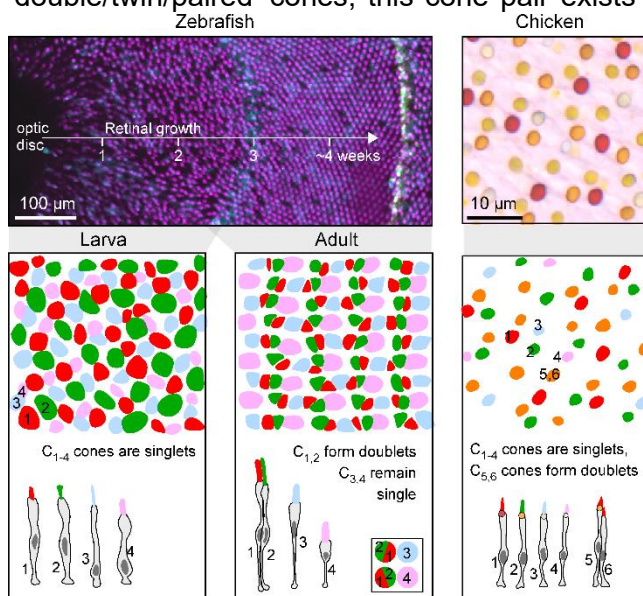


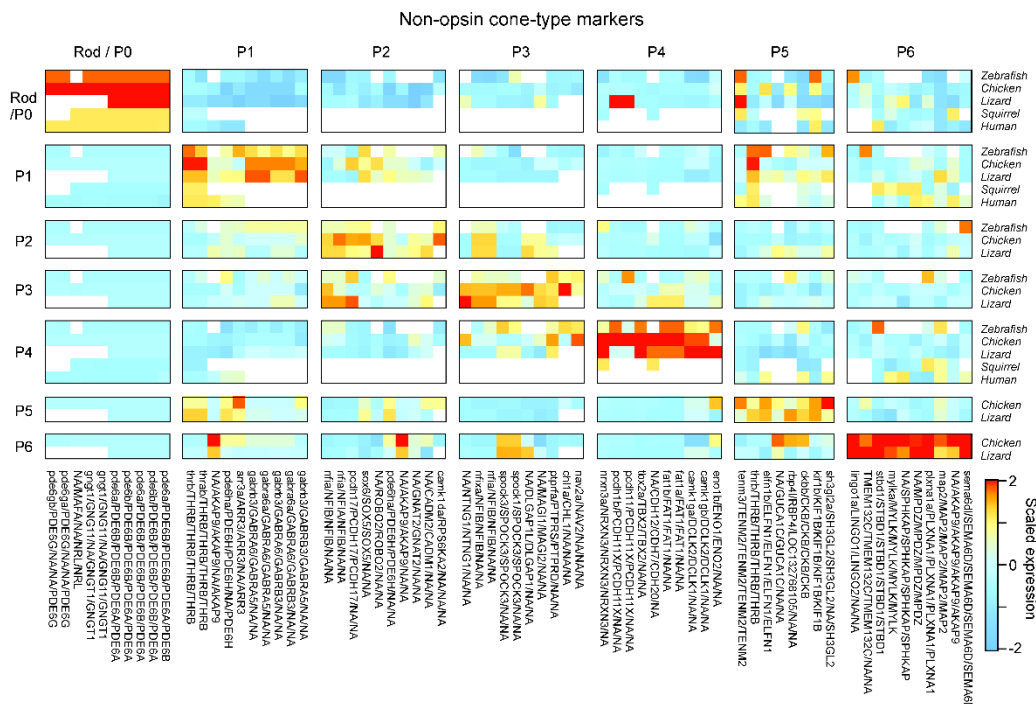
Figure: Left, Developmental transition of zebrafish cone patterning from a mosaic with low regularity to a highly regular row lattice (top, mod from Ref⁸⁵) and schematic summary of cone type arrangements at each stage (bottom, mod. from refs^{9,44,92}). Right, oil droplets in chicken retina show five independent cone-type mosaics (top) and schematic summary of cone types (bottom, both mod. from Ref⁷).

316 **Box 3. Naming yet-to-be-identified photoreceptor types.**

317 We recognize that the list of photoreceptor types presented in Fig. 1 is not exhaustive. For
 318 example, evidence suggests that the little skate, deep-sea fishes, marsupials, geckos, and
 319 snakes may possess unusual photoreceptor types that do not fall within any of the categories
 320 we propose. Furthermore, the photoreceptor types of only a minute fraction of the nearly
 321 67,000 extant vertebrate species have been studied in any detail. Thus, the present
 322 classification system is intended to be open-ended, permitting the addition of more
 323 photoreceptor types as they are characterized.

324 The goal of our definition is to include ‘extra’ types that exist *in parallel* to the ancestral ones
 325 (such as tetrapod double cones P5,6 or the amphibian ‘second rod’ P7), but to exclude those
 326 that represent within-retina variation (e.g. human P1-L versus P1-M).

327 More generally, the proposed definition of photoreceptors into ancestral types is centrally
 328 anchored in their distinct transcriptomic signatures¹⁷ across vertebrate ‘model’ species that
 329 are thought to be broadly representative for their clade: Humans (primates), squirrel (rodents),
 330 chicken (birds), anole lizard (non-avian reptiles), and zebrafish (teleosts). In these species,
 331 single cell transcriptomic data conforms with long standing insights into a photoreceptor’s
 332 morphological and functional properties (reviewed in Refs^{4,9,10,12,20}). Together, this wealth of
 333 data renders it unlikely that their cell-type definitions will need to be revised in the light of
 334 possible future evidence. We therefore posit that these species, and others where
 335 corresponding insights exist (e.g. Ref⁴³), can serve as a reference when defining cone types
 336 in other species.



343 generation of each cone type or for the expression of their unique set of genes; transcriptomic
344 datasets confirm that each cone type retains specific expression of these transcription factors
345 throughout their lifespan, likely actively controlling cone identity. Furthermore, evidence
346 suggests that transcription factors and their cognate binding sites throughout the genome are
347 interlocked and therefore highly resistant to evolutionary change. Thus, transcription factors
348 and *cis*-regulatory ‘grammars’ are often relatively fixed, ancestral features of a cell type which
349 are more persistent in evolution than the modules of ‘effector genes’ which they regulate, as
350 the latter are more directly subject to present-day adaptive pressures^{93,94}. Some examples of
351 deeply conserved photoreceptor transcription factors follow:

- 352 1. Generation of rods/P0 is dependent on Maf family transcription factors (e.g., NRL,
353 MAFA, MAFB) and NR2E3^{95,96}.
- 354 2. Generation of P1 depends on THRB, a subunit of the thyroid-hormone receptor⁹⁷⁻⁹⁹ as
355 well as the transcriptional co-factor SAMD7⁴⁷.
- 356 3. Generation of P2 depends on SIX6/SIX7^{100,101}.
- 357 4. Generation of P3 depends on FOXQ2; both P3 and FOXQ2 were lost in eutherian
358 mammals but retained in monotremes¹⁰²⁻¹⁰⁵.
- 359 5. Generation of P4 has been shown to depend on TBX2 in zebrafish^{48,106} and TBX2
360 expression is specific to P4 in transcriptomic datasets across vertebrates (zebrafish,
361 chicken, lizards, squirrels and primates).
- 362 6. Generation of P5 depends on THRB; and similar to P1, P5 also expresses SAMD7.
- 363 7. P6 expresses FOXQ2 and SKOR1, similar to P3 and P3,4, respectively.

364 Aside from the above, inferences about photoreceptor identity can nevertheless emerge from
365 limited data such as those used in classical definitions (Fig. 1) and reference to typical patterns
366 of photoreceptor properties across the vertebrate tree of life (reviewed e.g. in Refs^{4,13,20,24}).
367 For example:

- 368 1. Numerical abundance is usually $P1 \geq P2 > P3 \geq P4$. If present, P5,6 is usually
369 $P1 > P5/6 > P4$, except in birds, where the more typical pattern is $P5/6 > P1$.
- 370 2. If cone types are missing, the likely order of loss is $P2 = P3 > P4 > P1$. In non-eutherian
371 tetrapods, P5,6 is usually present. P5,6 are not known to occur individually.
- 372 3. Postsynaptic wiring appears to conform to ‘spectral blocks’ in the sense that
373 ‘intermediate’ cones, if present, do not tend to be skipped. For example, a bipolar cell
374 is unlikely to connect with P1 and P3 without also contacting P2. In this order, rods and
375 P5,6 appear to group with P1 (i.e. P0/P5,6-P1-P2-P3-P4). In birds, P6 appears to
376 additionally group with P3⁴⁵.
- 377 4. The spectral appearance of pigmented oil droplets, if present, generally correlates with
378 cone-type identity, with P1-P4 exhibiting long to short-wavelength filtering properties,
379 respectively, matching the spectral sensitivity of the corresponding opsins. P4 usually
380 has a clear oil droplet, devoid of light-absorbing carotenoid pigments. P5 tend to have
381 spectrally intermediate oil droplets and P6 tend to have either absent or minute
382 droplets, while frequently retaining carotenoid pigmentation in the mitochondrial
383 aggregates of the ellipsoid¹³.

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